

Quorum-Coupled Bacteriocin Release: Engineering a Champion

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Microorganisms use pheromones to monitor their own population density as well as to detect and interact with other microbial species in a process known as quorum sensing. For instance, bacteria can disrupt the pheromone signals of other competing bacteria, effectively preventing their proliferation. In a similar sense, we, the University of Calgary's iGEM 2008 WetWare team, have exploited the natural communication systems involving Autoinducer-1 (AI-1) from *Vibrio fischeri* and Autoinducer-2 (AI-2) from *Vibrio harveyi*, to create a model biosensor system in *Escherichia coli*. We have engineered the genetic circuits necessary for the production of these pheromones into two populations of *E. coli* (termed Bad guy #1 and Bad guy #2, as per their respective Autoinducer). In addition, our third population of *E. coli* (termed Champion cell) acts as a biosensor by receiving these signal inputs and subsequently initiating transcription of specific *E. coli*-targeted bacteriocins (*i.e.* colicins). The presence of AI-1 induces the Champion to produce a colicin to which Bad guy #1 is susceptible, but to which Bad guy #2 is resistant, and vice-versa for AI-2. An additional aspect of our system is the ability of the Champion cell to report the presence of each specific Bad guy by producing a specific fluorescent protein (*i.e.* either red or green fluorescent protein) in tandem with the specific colicin as determined by the presence of either AI-1 or AI-2. We constructed this system using the molecular cloning methods used in the undergraduate International Genetically Engineered Machine (iGEM) competition. While the *Vibrio fischeri* components of our system were obtained as standardized parts from the iGEM Registry, we cloned and standardized all parts of *Vibrio harveyi* AI-2 quorum sensing system. All of the standardized parts are flanked by specific restriction endonuclease sites (*i.e.* iGEM BioBricks), thereby allowing for a simple and iterative directional cloning strategy for the construction of the necessary genetic circuits.